

REVISION OF THE STRUCTURES OF RHAZICINE AND RHAZIMINE, TWO ALKALOIDS FROM *MELODINUS ACUTIFLORUS*

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Key Word Index—*Melodinus acutiflorus*; Apocynaceae; alkaloids; structure revision of rhazicine and rhazimine; relative configuration; crystal structure; spectral data.

Abstract—From the leaves of *Melodinus acutiflorus* the two alkaloids rhazicine and rhazimine were isolated and identified with known substances. However, based on extensive spectral and X-ray analysis their structures published earlier have to be revised.

INTRODUCTION

In the past, plants of the genus *Melodinus* (Apocynaceae, Carissae) have been investigated extensively for alkaloids. Some of these alkaloids are indole alkaloids, others are built up biogenetically from the same building blocks (tryptamine and secologanin) but contain a quinoline instead of an indole chromophore. To date, only nine members of this alkaloid type have been isolated [1]. These are meloscine [2], 16-*epi* meloscine [2], 16-*epi* meloscine-*N*-oxide [3], scandine [2], meloscandonine [4], 19-*epi* meloscandonine [5], scandomeline [5], 19-*epi* scandomeline [5] and lanceomigine [6]. Comparably derived quinoline alkaloids (e.g. quinine) are known from plants of the family of Rubiaceae. *M. acutiflorus* itself was not then screened for alkaloids.

RESULTS AND DISCUSSION

From the chloroform extract of the leaves of *M. acutiflorus* we have isolated the two alkaloids rhazicine (1) and its anhydro product rhazimine (2). During chromatography rhazicine converted to rhazimine by loss of one

mol. of water. By treatment of 1 with acetic anhydride-pyridine a mixture of 1 and 2 was formed. Structure determination was performed on the main product, rhazicine (1), crystallized from MeOH and it gave the result shown in the Fig. 1 and Scheme 1.

Because of the structural changes between 1 [$\text{HO}-\text{C}(2)-\text{O}-\text{C}(17)-\text{N}-$] and 2 [$\text{C}(2)=\text{O}-\text{C}(17)=\text{N}-$] which are in the neighbourhood of the chromophores, the UV spectra of 1 and 2 show significant differences (see Experimental). In the IR spectrum of 1 $\nu_{\text{C}=\text{O}}^{\text{CHCl}_3}$ appears at 1742 and in that of 2 at 1740 and 1732 cm^{-1} .

The mass spectrum (EI mode) of 1 is characterized by the base peak at m/z 122 and a significant peak for the loss of water (m/z 350). The corresponding ion is further decomposed to m/z 322. Indicated by m^* signals (linked scan technique) the latter is transformed to m/z 263 and 122. This principal fragmentation pattern is given in Scheme 2.

Using 2D-NMR methods we have been able to assign completely the ^1H and the ^{13}C spectra of 1. The assignment of the signals in 2 was done on the basis of that of 1 and of additional decoupling experiments (see Tables 1, 2 and 3). Comparison of the ^{13}C spectra of 1 and 2 reveals close similarity except for C(2) and C(17) which differ

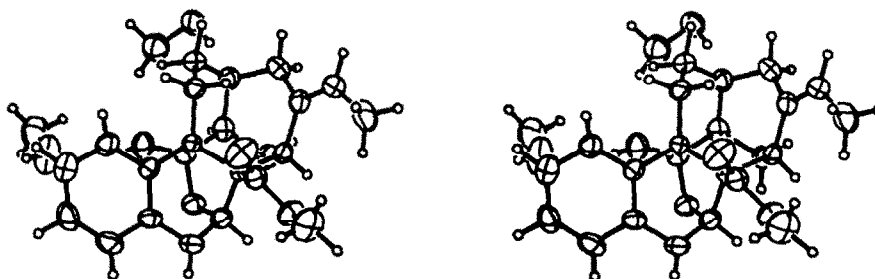
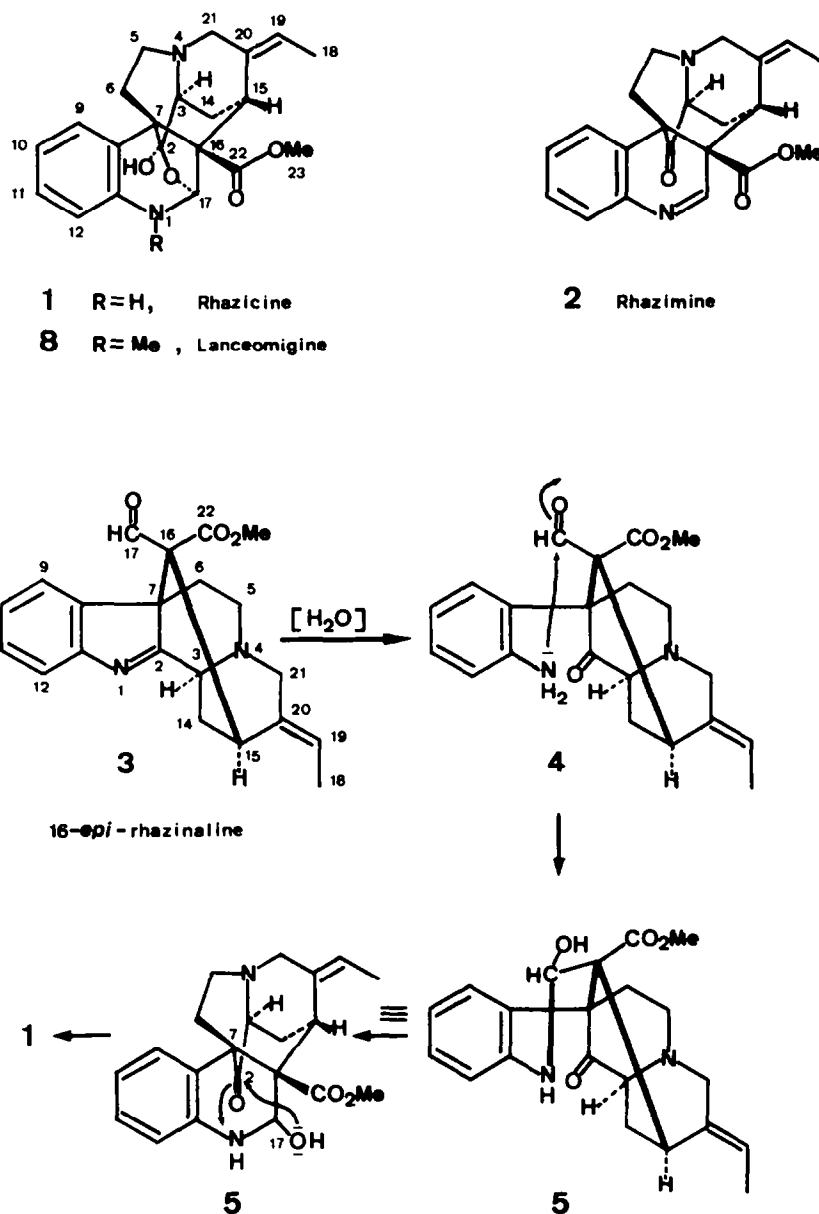


Fig. 1.



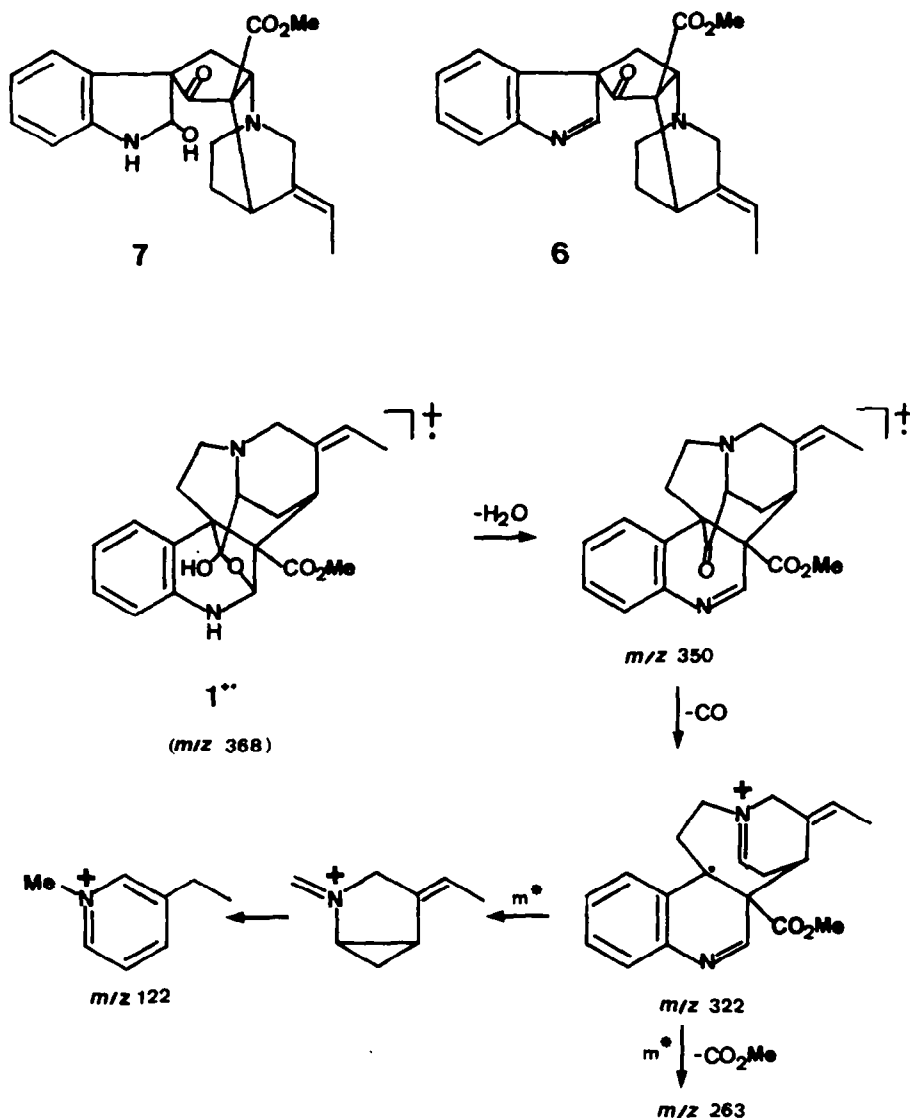
Scheme 1. Structure and possible biogenetic pathway of rhazicine (1).

from each other by more than 70 ppm. These findings are in full agreement with the structural differences between 1 and 2. On the basis of the extracted coupling constants for both compounds, the geometrical arrangement of 1 and 2 have to be similar and rhazimine must therefore possess structure 2.

The structure of rhazicine represents an interesting variation of *Melodinus* alkaloids. It can be assumed that the biogenetic pathway of rhazicine (1) begins with the known 16-*epi* rhazinaline (3) [7] which, under hydrolytic enzymatic conditions, is transformed to 4. This compound can undergo a ring closure reaction via the amina 5 to rhazicine (1) by acetalization of the ketone group (Scheme 1). X-ray structure determination gave the relative configuration of 1. If the postulated biogenetic pathway is correct and 16-*epi* rhazinaline is the precursor

of 1, its structure represents the absolute configuration. It was demonstrated that all indole alkaloids of type 3 have the same absolute configuration at C(15) [8].

The names rhazicine and rhazimine for the two alkaloids isolated from *M. acutiflorus* are the same as used for alkaloids isolated from *Rhazya stricta* Decaisne. The alkaloid sample of rhazimine (2) isolated from *R. stricta* [9] kindly provided by Atta-ur-Rahman and that of *M. acutiflorus* show the same behavior on TLC (silica gel: CHCl_3 -MeOH-HOAc, 8:9:1, R_f = 0.50; Et₂O-MeOH-aq. NH₄OH, 80:19:1, R_f = 0.47; EtOAc, R_f = 0.29), the same colour reaction with $\text{Ce}(\text{SO}_4)_2$ reagent, the same EI mass spectra and the same 400 MHz ¹H NMR spectra. Therefore, both compounds are identical. The structure of rhazimine given in [9] as 6 is incorrect and has to be changed to 2. Because of the



Scheme 2. Mass spectral fragmentation of rhazicine (1).

correlation of 2 and 1 (see above), the structure of rhazicine given in [10] as 7 must be changed to 1. We have to assume that 2-methoxy-1,2-dihydrorhazimine [11] has to be corrected, too.

An alkaloid called lanceomigine was isolated from *Alstonia lanceolata* V. Heurck et Müll. Arg. [12]. Its structure was given as *N*(1)-methyl-rhazicine (8). The structure determination was based on extensive spectral analysis and on an X-ray structure determination of a derivative [13]. Methylation ($H_2CO-HOAc-NaCNBH_3$) of rhazicine (1) gave a product which shows great similarity with the published spectral data of lanceomigine [6].

EXPERIMENTAL

1H NMR spectra were recorded at 400 and ^{13}C NMR at 50 MHz, 2D experiments at 400 MHz, using the $CHCl_3$ signal as

ref.; UV spectra were recorded in 95% EtOH, IR spectra in $CHCl_3$ and EIMS were measured at 70 eV.

Plant material. *M. acutiflorus* S. V. Müll. was collected near Brisbane/Australia.

Extraction and chromatography. Dried leaves (500 g) were extd $\times 4$ with 3% MeOH-HOAc at room temp. The MeOH extract was evapd ($< 40^\circ$), dil. with 0.25 N aq HCl and extd several times with Et_2O to remove the non-basic compounds. The aq phase was basified with Na_2CO_3 to pH 7, extd with Et_2O (AM1) and $CHCl_3$ (AM2), and continued to be basified to pH 9, extd with $CHCl_3$ (AM3). The organic soln was evapd ($< 40^\circ$). After chromatography on silica gel using $CHCl_3$ -MeOH- NH_4OH (varying ratios) we obtained from AM2 and AM3 600 mg of compound (1) and small amounts of (2).

Rhazicine (1). Crystallized from MeOH in colourless prisms, mp $122.6-124.4^\circ$; $[\alpha]_D^{22} + 69.8^\circ$ ($CHCl_3$, $c = 0.76$); pink colour with $Ce(SO_4)_2$. UV λ_{max} nm: 208 (log ϵ 4.16), 249 (3.70), 294 (3.22), λ_{min} nm: 228 (3.34), 272 (2.85); IR ν_{max} cm^{-1} : 3630, 3525,

Table 1. ^{13}C NMR spectral data (CDCl_3) of rhazicine (1) and rhazimine (2)

Carbon	1	2
2	111.12 s	214.51 s
3	57.40 d	61.17 d
5	48.31 t	51.65 t
6	23.10 t	30.39 t
7	49.72 s	53.14 s
8	126.19 s	137.43 s
9	127.29 d	128.36 d*
10	119.41 d	128.01 d*
11	128.44 d	128.84 d*
12	115.81 d	120.76 d
13	142.52 s	142.03 s
14	27.65 t	32.16 t
15	36.89 d	37.35 d
16	57.40 s	58.09 s
17	84.46 d	160.60 d
18	12.62 q	13.03 q
19	118.18 d	125.05 d*
20	141.08 s	137.43 s
21	54.14 t	53.04 t
22	170.91 s	168.42 s
23	51.69 q	52.16 q

*Interchangeable.

Table 2. ^1H NMR spectral data of rhazicine (1) (400 MHz)

Hydrogen	$\delta(\text{ppm})$	Multiplicity	$J(\text{Hz})$
1*	4.83	d	$J_{(1,17)} = 4.4$
3	3.56	dd†	$J_{(3,14a)} = 3.4, J_{(3,14\beta)} = 2.4$
5 α	3.56	dt	$J_{(5a,5\beta)} = J_{(5a,6\beta)} = 13.8, J_{(5a,6a)} = 3.8$
5 β	2.76	dd	$J_{(5\beta,5a)} = 13.8, J_{(5\beta,6\beta)} = 5.3$
6 α	2.23	dd	$J_{(6a,6\beta)} = 16.2, J_{(6a,5a)} = 3.8$
6 β	2.92	ddd	$J_{(6\beta,6a)} = 16.2, J_{(6\beta,5a)} = 13.8, J_{(6\beta,5\beta)} = 5.3$
9	7.30	dd	$J_{(9,10)} = 7.7, J_{(9,11)} = 1.1$
10	6.87	dt	$J_{(10,9)} = J_{(10,11)} = 7.7, J_{(10,12)} = 1.1$
11	7.11	dt	$J_{(11,10)} = J_{(11,12)} = 7.7, J_{(11,9)} = 1.1$
12	6.57	dd	$J_{(12,11)} = 7.7, J_{(12,10)} = 1.1$
14 α	2.04	ddd	$J_{(14a,14\beta)} = 13.9, J_{(14a,15)} = 1.9, J_{(14a,3)} = 3.4$
14 β	2.32	ddd	$J_{(14\beta,14a)} = 13.9, J_{(14\beta,3)} = 2.4, J_{(14\beta,15)} = 4.1$
15	3.62	ddd†	$J_{(15,19)} = 1, J_{(15,14a)} = 1.9, J_{(15,14\beta)} = 4.1$
17	4.91	d	$J_{(17,1)} = 4.4$
18(3H)	1.49	dd	$J_{(18,19)} = 7.1, J_{(18,21a)} = 2.5$
19	5.38	ddq	$J_{(19,18)} = 7.1, J_{(19,21a)} = 2.5, J_{(19,15)} = 1$
21 α	3.97	dquint	$J_{(21a,21\beta)} = 16.5, J_{(21a,19)} = J_{(21a,18)} = 2.5$
21 β	2.93	d	$J_{(21\beta,21a)} = 16.5$
23(3H)	3.57	s	
2(OH)*	3.03	s(br)	

The assignments of the α and β protons at C(14) and C(21) in 1 were made on the basis of homonuclear NOE experiments.

*With D_2O interchangeable.

†The multiplicity and the coupling constants of these signals were determined by indirect methods.

3415 (NH and OH bands), 1742 (ester band), 1725, 1609 (tetrahydroquinoline), 1476. ^1H NMR and ^{13}C NMR see Tables. CIMS (2-methylpropane): 369 ($[\text{M} + 1]^+$), 351 ($[\text{M} + 1 - 18]^+$). EIMS: m/z (rel. int.): 368 (8, $[\text{M}]^+$), 350 (15), 322 (16), 277 (6), 263 (12), 214 (11), 180 (10), 169 (10), 154 (14), 122 (100), 121 (21), 108 (11).

X-ray diffraction analysis of rhazicine-2 MeOH at room temp. Formula of the asymmetric unit $\text{C}_{21}\text{H}_{24}\text{N}_2\text{O}_4 \cdot 2 \text{ MeOH}$, space group $P2_12_12_1$; $a = 7.649(1)$, $b = 15.671(1)$, $c = 18.152(1)$ Å [from a least squares fit to the 2θ -values of 92 automatically centred reflections in the range $26 < |2\theta| < 33^\circ$, $\lambda(\text{MoK}\alpha) = 0.71069$ Å]. 2836 symmetry independent reflections within $\lambda^{-1} \sin \theta < 0.65$ Å were measured on a Nicolet R3 diffractometer using the ω -scan technique and were corrected for Lorentz and polarisation effects but not for absorption. The structure was solved by direct methods [14]. All H atoms were found by a difference density calculation and were freely refined with isotropic temperature factors, an exception being the H atoms in the methyl group of one MeOH molecule for which the riding model with $U = 0.2$ Å was used. All other atoms were freely refined with anisotropic temperature factors. The 2390 structure factors with $F > \sigma_F$ were used in a blocked cascade least squares process (ca 100 variables per block) [14] to refine the 396 variables to $R = 0.101$, $wR = 0.053$ for all 2836 reflections, $w = (\sigma_F^2 + 0.0004F^2)^{-1}$. The e.s.d.s. of the bond lengths range from 0.004 to 0.007 Å. The absolute structure was not determined. For atom coordinates and temperature factors see Table 4.

Table 3. ^1H NMR spectral data of rhazimine (2) (400 MHz)

Hydrogen	$\delta(\text{ppm})$	Multiplicity	$J(\text{Hz})$
3	3.91	<i>d</i>	$J_{(3,14\beta)} = 5.3$
5 α	3.78	<i>ddd</i>	$J_{(5\alpha,5\beta)} = 15.6$, $J_{(5\alpha,6\beta)} = 14.4$, $J_{(5\alpha,6\alpha)} = 5.8$
5 β	2.80	<i>dd</i>	$J_{(5\beta,5\alpha)} = 15.6$, $J_{(5\beta,6\beta)} = 3.5$
6 α	2.99	<i>dd</i>	$J_{(6\alpha,6\beta)} = 14.4$, $J_{(6\alpha,5\alpha)} = 5.8$
6 β	3.06	<i>dt</i>	$J_{(6\beta,6\alpha)} = J_{(6\beta,5\alpha)} = 14.4$, $J_{(6\beta,5\beta)} = 3.5$
9–12	7.4–7.3	<i>m</i>	
14 α	2.11	<i>dd</i>	$J_{(14\alpha,14\beta)} = 14.8$, $J_{(14\alpha,15)} = 2.5$
14 β	2.48	<i>ddd</i>	$J_{(14\beta,14\alpha)} = 14.8$, $J_{(14\beta,3)} = 5.3$, $J_{(14\beta,1)} = 2.7$
15	3.76	<i>dd</i> *	$J_{(15,14\alpha)} = 2.5$, $J_{(15,14\beta)} = 2.7$, $J_{(15,19)} = 2$
17	7.70	<i>s</i>	
18(3H)	1.59	<i>dd</i>	$J_{(18,19)} = 7.1$, $J_{(18,21\beta)} = 2.5$
19	5.58	<i>ddq</i>	$J_{(19,18)} = 7.1$, $J_{(19,21\beta)} = 2.5$, $J_{(19,15)} = 2$
21 α	4.12	<i>d quint</i>	$J_{(21\alpha,21\beta)} = 17.3$, $J_{(21\alpha,18)} = J_{(21\alpha,19)} = 2.5$
21 β	3.25	<i>d</i>	$J_{(21\beta,21\alpha)} = 17.3$
23(3H)	3.52	<i>s</i>	

* The multiplicity and the coupling constants of this signal were determined by indirect methods.

Table 4. Atom coordinates ($\times 10^4$) and temperature factors ($\text{\AA}^2 \times 10^3$) for rhazicine (1) dimethanol solvate

Atom	<i>x/a</i>	<i>y/b</i>	<i>z/c</i>	<i>U_{eq}</i> *
N(1)	3112(4)	6253(2)	3789(2)	42(1)
C(2)	5444(5)	4988(2)	2763(2)	34(1)
O(2)	6847(3)	5330(2)	2391(1)	47(1)
C(3)	5020(5)	4114(2)	2407(2)	38(1)
N(4)	6148(4)	3408(2)	2698(2)	40(1)
C(5)	7540(4)	3707(2)	3196(2)	40(1)
C(6)	6864(4)	4186(2)	3858(2)	35(1)
C(7)	5604(4)	4897(2)	3615(2)	28(1)
C(8)	6122(4)	5784(2)	3914(2)	31(1)
C(9)	7824(5)	6000(2)	4080(2)	39(1)
C(10)	8307(5)	6812(3)	4302(2)	48(1)
C(11)	7062(6)	7425(2)	4365(2)	51(1)
C(12)	5327(5)	7253(2)	4192(2)	47(1)
C(13)	4869(5)	6434(2)	3961(2)	35(1)
C(14)	3096(5)	3939(2)	2551(2)	39(1)
C(15)	2893(4)	3876(2)	3396(2)	33(1)
C(16)	3612(4)	4721(2)	3760(2)	28(1)
C(17)	2905(5)	5501(2)	3349(2)	37(1)
O(17)	3948(3)	5550(1)	2683(1)	38(1)
C(18)	1831(8)	2698(4)	4711(3)	63(2)
C(19)	3251(5)	2550(2)	4157(2)	45(1)
C(20)	3735(5)	3045(2)	3609(2)	37(1)
C(21)	5093(5)	2725(2)	3057(2)	46(1)
C(22)	3172(5)	4749(2)	4575(2)	38(1)
O(22)	4184(4)	4690(2)	5070(1)	59(1)
C(23)	839(7)	4907(4)	5421(3)	69(2)
O(23)	1431(3)	4817(2)	4672(1)	47(1)
O(M1)	8579(4)	2883(2)	1685(2)	55(1)
C(M1)	9019(8)	3627(4)	1249(3)	64(2)
O(M2)	9202(4)	6664(2)	2517(2)	86(1)
C(M2)	10144(6)	6438(3)	1868(2)	75(2)

* Equivalent isotropic *U* defined as one third of the trace of the orthogonalized *U* tensor.

Rhazimine (2). Crystallized from MeOH, mp 170.5° (dec.); $[\alpha]_D^{25} = +252.0^\circ$ (CHCl_3 , *c* 0.23); pink colour with $\text{Ce}(\text{SO}_4)_2$; UV λ_{max} nm: 202 nm (log *e* 4.30), 218 (4.22), 260 (3.63), 288 (sh), λ_{min} nm: 214 (4.21), 251 (3.62); IR λ_{max} cm^{-1} : 1740 (ester), 1732 (ketone), 1628; ^1H NMR and ^{13}C NMR see Tables. EIMS *m/z* (rel. int.): 350 (4, $[\text{M}]^+$), 322 (5), 307 (1), 291 (1), 263 (2), 214 (4), 122 (100), 108 (9), 93 (6), 77 (5).

Rhazimine (2) from rhazicine (1). Treatment of a sample of (1) with Ac_2O -pyridine gave a mixture of rhazimine (2) as main product and starting material; identification by TLC. During chromatography (silica gel; CHCl_3 -MeOH- NH_4OH) of (1) rhazimine was formed in small quantities.

Conversion of rhazicine (1) to lanceomigine (8). 1 (10 mg) was dissolved in a soln of HCHO (5 ml, 40% in H_2O) and HOAc (5 drops). Afterwards at 20°, NaCNBH_3 was added slowly under stirring. After 15 min the reaction mixt was poured into ice- H_2O , basified with satd aq NaHCO_3 , extd with CHCl_3 , dried and evapd. Chromatography of the residue gave 8. Pink colour with $\text{Ce}(\text{SO}_4)_2$. ^1H NMR: 5.48 (1H, *q*, H-19), 4.82 (1H, *s*, H-17), 3.57 (3H, *s*, O- CH_3), 2.98 (3H, *s*, N- CH_3), 1.51 [3H, *dd*, CH_3 (18)]. EIMS *m/z* (rel. int.): 382 (40, $[\text{M}]^+$), 367, 338, 337, 323, 279, 222, 207, 194, 167, 158, 157, 122 (100), 121.

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